

Human Gastric Organoid Kit

Introduction

This kit is a comprehensive system for the establishment and maintenance of human gastric organoids. This kit contains a complete set of reagents for tissue preservation, organoid construction, organoid passage, and cryopreservation. This kit is ideal for organoid culture and does not require additional reagents other than Matrigel. Organoids cultured with this kit have a gastric epithelium-like structure and are ideal models for gastric physiology and pathology studies.

Components and Storage

Components	Size	5 Assays	Storage
Human Gastric Organoid Growth Medium		100 mL	-20°C
Gastrointestinal Tissue Dissociation Solution		50 mL	4°C
Organoid Dissociation Solution		40 mL	-20°C
Organoid Cryopreservation Medium		40 mL	4°C
Tissue Storage Solution		100 mL	-20°C
Organoid Wash Buffer		500 mL	-20°C
Shipping: Dry ice		Shelf life: 6 months	

Materials Required but Not Included

Products	Catalog number
GFR Basement Membrane Matrix (Phenol Red-Free)	EM1001
100 µm cell strainer	-
24-well cell culture plate	-
15 mL centrifuge tubes	-
1.5 mL EP tubes	-

Protocol

1. Preparation before use:

1. Thaw GFR Basement Membrane Matrix (Phenol Red-Free) at 4°C overnight in advance.

2. Thaw components stored at -20°C in advance, and aliquot the Human Gastric Organoid Growth Medium to avoid repeated freeze-thaw cycles.

2. Construction of organoids from primary tissues

1. Tissue isolation

- 1) Isolate tissue in accordance with local ethics regulations. Under sterile conditions, isolate tissue, wash tissue with saline several times, and then place in a conical tube containing Tissue Storage Solution. Keep tissue at 4°C until start the experiment.

***Note:** For detailed protocol, please refer to the manual of Tissue Storage Solution (K2842).

2) Processing Based on Tissue Fragment Size.

A. Small fragments (approx. 5-8 mm)

- i. Transfer the pieces to a dish and further cut it into $2\text{-}4\text{ mm}^3$ pieces with surgical scissors.
- ii. Add 5-10 mL of Organoid Wash Buffer to pipette and wash repeatedly.
- iii. Remove the Organoid Wash Buffer.
- iv. Repeat steps ii-iii for a total of 5-10 wash of tissue.
- v. At this point, 1–2 tissue pieces can be collected for in situ analysis.
- vi. Transfer the pieces to a 1.5 mL centrifuge tube and further cut them into 0.5 mm^2 pieces with surgical scissors.
- vii. Transfer the pieces to a 15 mL centrifuge tube.
- viii. Add 6 mL of Gastrointestinal Tissue Dissociation Solution, seal the tube with parafilm, and place in 4°C to digest.
- ix. Every 5 min, vortex for 30 s, and aspirate 30 μL of suspension and observe under the microscope.
- x. The digestion time should be controlled within 40 min, and the digestion can be stopped when observing 10-100 cell clumps around 100 μm in diameter under the microscope.

***Note:** Over-digestion (e.g., single cell) can affect subsequent organoid construction and reduce viability.

- xi. Filter the suspension by a 100 μm cell strainer and supplement with Organoid Wash Buffer to 26 mL.
- xii. Evenly divide the filtered suspension into two 15 mL centrifuge tubes and then centrifuge at 300 g or 1500 rpm for 5 min. Discard the supernatant.
- xiii. If observing a red precipitate, add 2 mL of Red Blood Cell Lysis Buffer (K1169) to resuspend the precipitate, and then let it stand at room temperature for 3 min.

***Note:** If the tissue has been harvested for more than 48 h, it is not recommended to use Red Blood Cell Lysis Buffer.

- xiv. Add 10 mL of Organoid Wash Buffer. Centrifuge at 300 g or 1500 rpm for 5 min and discard the supernatant.

***Note:** After centrifugation, remove the supernatant as much as possible. Because the concentration of Matrigel needs to be above 70% to make structural stability.

B. Large fragments (>1 cm)

- i. Transfer the pieces to a culture dish. Then cut them into 1 cm tissue pieces.
- ii. Add 3–8 mL of Organoid Wash Buffer. Use forceps to fix one end of the tissue and carefully scrape off the upper layer of the mucosa lining the tissue with a clean slide.
- iii. Remove the Organoid Wash Buffer and transfer the tissue pieces to a 15 mL centrifuge tube.
- iv. Add 5–8 mL of Organoid Wash Buffer.
- v. Vortex for 30 s, let the centrifuge tube stand for 1 min, and discard the supernatant.
- vi. Repeat steps iv-v for a total of 5-8 wash of tissue.
- vii. At this point, 1–2 tissue pieces can be collected for in situ analysis.
- viii. Add 6-8 mL of Gastrointestinal Tissue Dissociation Solution, seal the tube with parafilm, and place in 4°C to digest.
- ix. Every 5 min, vortex for 30 s, and aspirate 30 µL of suspension and observe under the microscope.
- x. After 30-40 min of digestion, remove the centrifuge tube from the refrigerator and transfer the contents to a culture dish.
- xi. Use clean forceps to hold one end of the tissue. Take a clean glass slide and scrape the tissue surface repeatedly at a 45° angle. After scraping all the tissue, place the culture dish under a microscope for examination.
- xii. Stop the digestion when a large number of gastric gland structures are observed.

***Note:** Over-digestion (e.g., single cell) can affect subsequent organoid construction and reduce viability.

- xiii. Collect the supernatant and transfer it to a 15 mL centrifuge tube. Top up with Organoid Wash Buffer to 11 mL. Centrifuge at 300 g or 1500 rpm for 5 min and discard the supernatant.

***Note:** After centrifugation, remove the supernatant as much as possible. Because the concentration of Matrigel needs to be above 70% to make structural stability.

2. Organoid culture

- 1) Mix the pellet and GFR Basement Membrane Matrix (Phenol Red-Free) on ice at a ratio of 1:10. Thaw GFR Basement Membrane Matrix (Phenol Red-Free) at 4°C overnight in advance.

***Note:** If air bubbles appear during the blowing process, absorb the Matrigel below the bubbles, and then vigorously tap the table with the tube to break the bubbles.

- 2) Using a 24-well plate as an example, drop the Matrigel/Cell mixture evenly in the center of the well, 20-60 µL per drop.
- 3) Place the plate in a 37°C incubator to set the Matrigel domes.
- 4) After 15 min, add 500 µL of room temperature Human Gastric Organoid Growth Medium to each well by pipetting the medium gently down the sidewall. Do not pipette the medium onto the domes directly.

***Note:** Equilibrate Human Gastric Organoid Growth Medium at room temperature for 15 min in advance.

- 5) Incubate the plate in a 37°C, 5% CO₂ incubator. Exchange Human Gastric Organoid Growth Medium every 2-4 days.

***Note:**

- a) The time for exchanging medium depends on the experimental situation. If the cell density is high, the medium turns yellow after one day, and it is necessary to change the medium daily and dilute the cell density as soon as possible.
- b) If microbial contamination occurs during the incubation process, add 1 mL of 3.5 mM NaOH solution to the contaminated wells and replace with Human Gastric Organoid Growth Medium after 4 h.

3. Organoid passage

1. Organoid harvest

- 1) Discard the medium and slowly add pre-chilled Organoid Wash Buffer to the wells (PBS containing 1% antibiotics can also be used).
- 2) Pipet Matrigel/Organoid/Medium mixture up and down to release organoid from the Matrigel.

***Note:** If finding that organoids are attached to the wall, scrape the organoid off with a pipette tip.

- 3) Transfer the mixture to a new 15 mL tube and add Organoid Wash Buffer to make up the volume to 12 mL.
- 4) Place the tubes in -20°C for 6 min or 4°C for 30 min.

***Note:** Brief periods of low temperature do not affect the activity of organoids, and Matrigel can also be removed by taking advantage of the properties of Matrigel dissolving at low temperatures.

- 5) Centrifuge at 300 g or 1500 rpm for 5 min, and discard the supernatant.

2. Organoid dissociation

Organoids are generally passaged through a combination of mechanical and enzymatic dissociation. Some organoids may only require mechanical dissociation, while others may only require enzymatic dissociation.

Some may require both methods. Not to reduce organoids to single cells during passaging.

Mechanical dissociation method

- 1) Suspend pellet in 1-2 mL of Organoid Wash Buffer. Pipet the suspension up and down.
- 2) When observing the organoids reach 40-200 μm in diameter, stop the digestion.
- 3) Add Organoid Wash Buffer to a total volume of 12 mL.
- 4) Centrifuge at 300 g or 1500 rpm for 5 min. Discard the supernatant.

Enzymatic dissociation method

- 1) Suspend pellet in 1-2 mL of room temperature Organoid Dissociation Solution. Pipet the suspension up and down.

***Note:** Equilibrate Organoid Dissociation Solution at room temperature in advance.

- 2) Every 2 min, observe the size of organoids. When the organoids reach 40-200 μm in diameter, stop the digestion. It is recommended that the digestion time should not exceed 6 min.
- 3) Add Organoid Wash Buffer to 12 mL.
- 4) Centrifuge at 300 g or 1500 rpm for 5 min. Discard the supernatant.

3. Organoid passage

- 1) Mix the pellet and GFR Basement Membrane Matrix (Phenol Red-Free) on ice at a ratio of 1:15. Thaw GFR Basement Membrane Matrix (Phenol Red-Free) at 4°C overnight in advance.

***Note:** If air bubbles appear during the blowing process, absorb the Matrigel below the bubbles, and then vigorously tap the table with the tube to break the bubbles.

- 2) Using a 24-well plate as an example, drop the Matrigel/Organoids mixture evenly in the center of the well, 20-60 μL per drop.
- 3) Place the plate in a 37°C incubator to set the Matrigel domes.
- 4) After 15 min, add 500 μL of room temperature Human Gastric Organoid Growth Medium to each well by pipetting the medium gently down the sidewall. Do not pipette the medium onto the domes directly.

***Note:** Equilibrate Human Gastric Organoid Growth Medium at room temperature for 15 min in advance.

- 5) Incubate the plate in a 37°C, 5% CO₂ incubator. Exchange Human Gastric Organoid Growth Medium every 2-4 days.

***Note:**

- a) The time for exchanging medium depends on the experimental situation. If the cell density is high, the medium turns yellow after one day, and it is necessary to change the medium daily and dilute the cell density as soon as possible.

- b) If microbial contamination occurs during the incubation process, add 1 mL of 3.5 mM NaOH solution to the contaminated wells and replace with Human Gastric Organoid Growth Medium after 4 h.

4. Organoid cryopreservation

Cryopreservation must be performed when the organoids are in optimal condition. Fast-growing organoids (up to 300-500 µm in diameter) can be cryopreserved when they reach 150-200 µm in diameter. Slow-growing organoids (up to 100-300 µm in diameter) can be cryopreserved when they reach 100-150 µm in diameter.

During the cryopreservation, gently pipetting to ensure the integrity of the organoids.

1. Organoid harvest

- 1) Discard the medium and slowly add pre-chilled Organoid Wash Buffer to the wells (PBS containing 1% antibiotics can also be used).
- 1) Pipet Matrigel/Organoid/Medium mixture up and down to release organoid from the Matrigel.

***Note:** If finding that organoids are attached to the wall, scrape the organoid off with a pipette tip.

- 2) Transfer the mixture to a new 15 mL tube and add Organoid Wash Buffer to make up the volume to 12 mL.
- 3) Place the tubes in -20°C for 6 min or 4°C for 30 min.

***Note:** Brief periods of low temperature do not affect the activity of organoids, and Matrigel can also be removed by taking advantage of the properties of Matrigel dissolving at low temperatures.

- 4) Centrifuge at 300 g or 1500 rpm for 5 min, and discard the supernatant.

2. Organoid cryopreservation

- 1) Depending on the pellet volume, mix the pellet and Organoid Cryopreservation Medium at a ratio of 1:10.
- 2) After mixing, aliquot the organoids into cryopreservation vials, and perform cryopreservation by programmed cooling.

***Note:** Cryopreserved organoids can be stored at -80°C for one month, and for long-term storage, store the vials in a liquid nitrogen tank.

3. Organoid resuscitation

- 1) Equilibrate Organoid Wash Buffer to room temperature in advance. Add 2 mL of room temperature Organoid Wash Buffer in a 15 mL centrifuge tube. 2% serum can be added to the Organoid Wash Buffer.
- 2) Remove the vials from the liquid nitrogen tank and thaw them in a 37°C water bath for 1-2 min.
- 3) Slowly transfer the thawed organoid suspension to the 15 mL centrifuge tube containing Organoid Wash Buffer. Add another 8 mL of Organoid Wash Buffer.

- 4) Centrifuge at 300 g or 1500 rpm for 5 min. Discard the supernatant.
- 5) Add 2 mL of room temperature Organoid Wash Buffer (without serum) and gently resuspend the organoids.
- 6) Add 8 mL of Organoid Wash Buffer.
- 7) Centrifuge at 300 g or 1500 rpm for 5 min. Discard the supernatant.

4. Organoid culture

- 1) Mix the pellet and GFR Basement Membrane Matrix (Phenol Red-Free) on ice at a ratio of 1:10. Thaw GFR Basement Membrane Matrix (Phenol Red-Free) at 4°C overnight in advance.

***Note:** If air bubbles appear during the blowing process, absorb the Matrigel below the bubbles, and then vigorously tap the table with the tube to break the bubbles.

- 2) Using a 24-well plate as an example, drop the Matrigel-Cell mixture evenly in the center of the well, 20-60 µL per drop.
- 3) Place the plate in a 37°C incubator to set the Matrigel domes.
- 4) After 15 min, add 500 µL of room temperature Human Gastric Organoid Growth Medium to each well by pipetting the medium gently down the sidewall. Do not pipette the medium onto the domes directly.

***Note:** Equilibrate Human Gastric Organoid Growth Medium at room temperature for 15 min in advance.

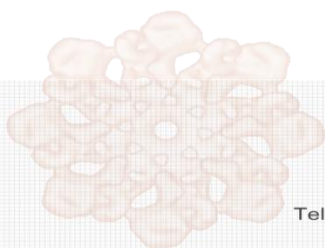
- 5) Incubate the plate in a 37°C, 5% CO₂ incubator. Exchange Human Gastric Organoid Growth Medium every 2-4 days.

***Note:**

- a) The time for exchanging medium depends on the experimental situation. If the cell density is high, the medium turns yellow after one day, and it is necessary to change the medium daily and dilute the cell density as soon as possible.
- b) If microbial contamination occurs during the incubation process, add 1 mL of 3.5 mM NaOH solution to the contaminated wells and replace with Human Gastric Organoid Growth Medium after 4 h.

Note

1. For your safety and health, please wear lab coats and gloves during the experiment.
2. For research use only. Not to be used in clinical diagnostic or clinical trials.



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