

## Celestine Blue Hematoxylin Stain Kit

### Introduction

Plain alum hematoxylin has a major drawback after nuclear staining in that it is relatively sensitive to subsequent acidic dyes, which is common in Van Gieson staining and Masson trichrome staining. During Van Gieson staining, the use of picric acid-fuchsin often causes most of the hematoxylin color to be washed out, making the nuclei barely distinguishable. To address this issue, iron hematoxylin can be used in combination with Celestine Blue, as Celestine Blue is acid-resistant. Moreover, the ferric salts in the Celestine Blue solution can enhance the binding force of alum hematoxylin to the nucleus, resulting in deeper nuclear staining.

### Components and Storage

Components	Size	50 mL	100 mL	Storage
	Reagent (A): Mayer Hematoxylin dye solution		50 mL	100 mL
Reagent (B): Celestine Blue solution		50 mL	100 mL	4°C protected from light
Shipping: Blue ice		Shelf life: 12 months		

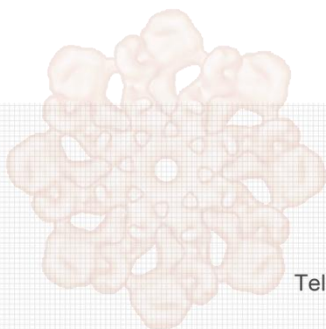
### Protocol

1. Deparaffinize the paraffin sections and hydrate through a graded ethanol series to distilled water.
2. Immerse in Celestine Blue solution for 2-3 min, then rinse with distilled water for 5-10 sec.
3. Add Mayer Hematoxylin dye solution for 2-3 min. Blue with tap water for 10 min.
4. Proceed with subsequent operations according to experimental requirements.

### Note

1. Staining intensity diminishes over time; prolong staining duration as needed. Increase staining time for tissues fixed in unbuffered formalin or decalcified tissues. Decrease staining time for frozen sections.
2. Alternatively, treat with bluing solution for 2-3 min to replace the tap water bluing step. Bluing solutions include 0.2-1% Ammonia Water, Scott's Bluing Solution, or 0.1-1% Lithium Carbonate Solution.
3. For your safety and health, please wear lab coats and gloves during the experiment.

4. For research use only. Not to be used in clinical diagnostic or clinical trials.



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