

Product Information

Neuraminidase Activity Fluorometric Assay Kit

I. Kit Contents:

Components	K2230-100 100 assays	Cap Color	Part Number
NA Assay Buffer	30 ml	WM	K2230-C-1
NA Probe (in DMSO, anhydrous)	200 µl	Red	K2230-C-2
NA Substrate (Lyophilized)	1 vial	Blue	K2230-C-3
NA Enzyme Mix I (Lyophilized)	1 vial	Green	K2230-C-4
NA Enzyme Mix II (Lyophilized)	1 vial	Purple	K2230-C-5
Galactose Standard (100 nmol/µl)	100 µl	Yellow	K2230-C-6
NA Positive Control	100 µl	Orange	K2230-C-7

II. Introduction:

Neuraminidase (NA) is an enzyme that catalyzes the hydrolysis of terminal sialic acid residues from the newly formed virions. NA plays an important role in the replication of influenza virus and the invasion of target cells. NA is also involved in the prevention of self-aggregation of virus and the elution of progeny viruses from infected cells. Hence, NA is an important target for the prevention of the spread of influenza virus.

The Neuraminidase Activity Fluorometric Assay Kit provides a sensitive, simple, fast and convenient way for detection of NA activity in various biological samples based on fluorometric method (Ex/Em = 530/590 nm). In the assay, NA Probe is used to detect the neuraminidase activity. The assay is suitable for high-throughput adaptable assay. The kit can detect as low as 2.0 mU/ml NA activity in a variety of samples.

III. Application:

Measurement of NA activity in various tissues/cells extracts or serum sample infected with influenza virus.

IV. Sample Type:

Animal tissues: liver, brain, kidney etc.

Cell culture: Adherent or suspension cells.

Serum.

V. User Supplied Reagents and Equipment:

96-well plate with flat bottom, preferably black plate.

Multi-well spectrophotometer with fluorescence.

VI. Storage and Handling:

Store kit at -20°C, protected from light. Briefly centrifuge small vials prior to opening. Read the entire protocol before performing the assay.

VII. Reagent Preparation and Storage Conditions:

NA Assay Buffer: Warm Assay Buffer to room temperature before use. Store at -20°C. Use within two months.

NA Probe: Warm to room temperature before use. Store at -20°C. Use within two months.

NA Substrate: Reconstitute with 110 μ l dH₂O. Store at -20°C. Use within two months.

NA Enzyme Mix I: Reconstitute with 220 μ l NA Assay Buffer. Aliquot and store at -20°C. Stable for two months.

NA Enzyme Mix II: Reconstitute with 220 μ l NA Assay Buffer. Aliquot and store at -20°C. Stable for two months.

NA Positive Control: Aliquot and store at -20°C. Use within two months.

VIII. Neuraminidase Assay Protocol:

1. Sample Preparation: Homogenize 10 mg of sample (wet weight or cell pellet) in 100 μ l NA Assay Buffer. Centrifuge at 10,000 x g for 5 min. at 4°C. Collect the supernatant. Add 2 - 10 μ l of supernatant or serum into a 96-well plate and adjust the volume to 50 μ l with NA Assay Buffer. Add 10 μ l of NA Positive Control into desired wells(s) and adjust the volume to 50 μ l with NA Assay Buffer.

Notes:

a. For unknown samples, we suggest testing several doses to ensure the readings are within the Standard Curve range.
 b. For samples having background, prepare parallel well(s) containing the same amount of sample as in the test well (sample background control). Adjust the volume to 50 μ l with NA Assay Buffer.

c. 1% or higher Triton X-100 concentration interferes with the assay.

2. Standard Curve Preparation: Dilute Galactose Standard to 1 nmol/ μ l by adding 10 μ l of 100 nmol/ μ l Galactose Standard into 990 μ l of NA Assay Buffer, mix well. Dilute the Standard further to 0.1 nmol/ μ l by adding 20 μ l of 1 nmol/ μ l Standard to 180 μ l of NA Assay Buffer and mix. Add 0, 1, 2, 3, 4 and 5 μ l of 0.1 nmol/ μ l Galactose Standard into a series of wells in 96-well plate to generate 0, 0.1, 0.2, 0.3, 0.4 and 0.5 nmol/well of Galactose Standard. Adjust the volume to 50 μ l/well with NA Assay Buffer.

We recommend using black plate for the assay.

3. Reaction Mix: Mix enough reagents for the number of assays (samples, Standards & Positive Control) to be performed. For each well, prepare 50 μ l Reaction Mix containing:

	Reaction Mix	Background Control Mix
NA Assay Buffer	44.5 μ l	45.5 μ l
NA Enzyme Mix I	2 μ l	2 μ l
NA Enzyme Mix II	2 μ l	2 μ l
NA Substrate	1 μ l	---
NA Probe	0.5 μ l	0.5 μ l

Add 50 μ l of Reaction Mix to each well containing the Standards, Positive Control and samples. Mix well.

For samples having background, add 50 μ l of Background Control Mix to sample background control well(s). Mix well.

4. Measurement: Incubate for 30 min. at 37°C and measure fluorescence (Ex/Em = 535/590 nm) kinetically.

Note: Incubation time depends on the NA Activity in the samples. Choose two time points (T1 and T2) in the linear range (fluorescence values A1 and A2 respectively) to calculate the NA activity of the samples. The Standard Curve can be read in end point mode (i.e. at the end of incubation time).

5. Calculations: Subtract 0 Standard reading from all readings. Plot the Galactose Standard Curve. If sample background control reading is significant, subtract background control reading from sample readings. Calculate the NA activity of the test sample: Δ RFU = A2 - A1. Apply Δ RFU to the Standard Curve to get B nmol of Galactose generated by NA during the reaction time (Δ T = T2 - T1).

Sample Neuraminidase Activity = $B / (\Delta T \times V) \times D = \text{nmol/min/ml} = \text{mU/ml}$

Where: B is the Galactose amount from the Standard Curve (nmol).

Δ T is the reaction time (min.).

V is the sample volume added into the reaction well (ml).

D is the sample dilution factor.

Unit Definition: One unit of Neuraminidase activity is the amount of enzyme that generates 1.0 μ mol of Galactose per min. at pH 7.4 at 37°C.

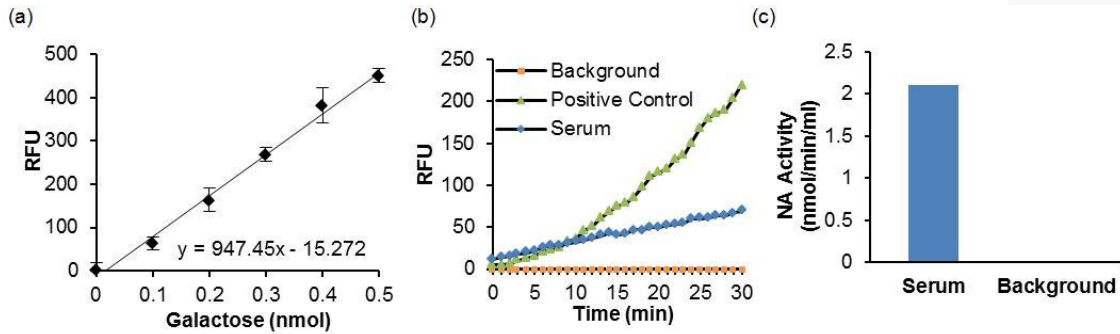


Figure: (a) Galactose Standard Curve. (b) NA activity in normal human serum (1 μ l) & Positive Control (1 μ l). (c) Calculated activity of serum. Assays were performed following the kit protocol.

For research use only! Not to be used in humans.

Our promise

If the product does not perform as described on this datasheet, we will offer a refund or replacement. For more details, please visit <http://www.apexbt.com/> or contact our technical team.

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