

## Product Information

### HDL and LDL/VLDL Quantification Colorimetric/Fluorometric Kit

#### I. Kit Contents:

Components	K2122-100 100 assays	Cap Color	Part Number
Cholesterol Assay Buffer	25 ml	WM	K2122-C-1
2X LDL/VLDL Precipitation Buffer	10 ml	NM	K2122-C-2
Cholesterol Probe (in DMSO, anhydrous)	200 µl	Red	K2122-C-3
Enzyme Mix (Lyophilized)	1 vial	Green	K2122-C-4
Cholesterol Esterase (Lyophilized)	1 vial	Blue	K2122-C-5
Cholesterol Standard (2 µg/µl)	100 µl	Yellow	B1702

#### II. Introduction:

High-density lipoprotein (HDL), low-density lipoprotein (LDL) and very low-density lipoprotein (VLDL) are three of the five major groups of lipoproteins and transport lipids within the water around cells. High levels of LDL and low levels of HDL are related to an increased risk of cardiovascular diseases. Regulation of LDL and HDL play critical roles in various disease developments.

The HDL and LDL/VLDL Quantification Colorimetric/Fluorometric Kit provides a sensitive, fast and convenient way for detection of HDL and LDL/VLDL in plasma and serum samples based on a convenient separation of HDL from LDL and VLDL, and colorimetric and fluorometric method. In the assay, cholesterol oxidase specifically recognizes free cholesterol and generates products which react with probe to yield fluorescence (Ex/Em = 538/587 nm) and color (570 nm). Cholesteryl ester can be hydrolyzed into free cholesterol by cholesterol esterase, therefore, cholesterol ester and free cholesterol can be separately detected in the absence and presence of cholesterol esterase in the reactions.

#### III. Reagent Preparation:

**Cholesterol Probe:** Ready to use as supplied. Warm to room temperature to thaw the DMSO solution. Store at  $-20^{\circ}\text{C}$ , protect from light. Use within two months.

**Cholesterol Esterase:** Dissolve in 220 µl Cholesterol Assay Buffer prior to use. Aliquot and store at  $-20^{\circ}\text{C}$ . Use within two months.

**Enzyme Mix:** Dissolve in 220 µl Cholesterol Assay Buffer prior to use. Aliquot and store at  $-20^{\circ}\text{C}$ . Use within two months.

#### IV. Cholesterol Assay Protocol:

1. Separation of HDL and LDL/VLDL: Mix 100 µl of 2X Precipitation Buffer with 100 µl of serum sample in microcentrifuge tubes. Incubate 10 min at room temperature, centrifuge at  $2000 \times g$  (5000 rpm on bench-top microcentrifuge) for 10 min. Transfer the supernatant into new labeled tubes. This is the HDL fraction. The precipitates are the LDL/VLDL fraction. If you want to measure the LDL/VLDL level, the precipitate should be spun again, and the trace amount of HDL supernatant carefully removed. Resuspend the precipitate in 200 µl PBS (not provided). This is the LDL/VLDL fraction.

Note

A: If the supernatant is cloudy, the sample should be re-centrifuged. If the sample remains cloudy, dilute the sample 1:1 with PBS, and repeat the separation procedure. Multiply final results by two (2) due to the dilution with the 2X Precipitation Buffer.

B: Precipitation time and temperature do not affect the results significantly.

2. Standard Curve and Sample Preparations: Dilute the Cholesterol Standard to 0.25  $\mu\text{g}/\mu\text{l}$  by adding 20  $\mu\text{l}$  of the Cholesterol Standard to 140  $\mu\text{l}$  of Cholesterol Assay Buffer, mix well. Add 0, 4, 8, 12, 16, 20  $\mu\text{l}$  into a series of wells in a 96-well plate. Adjust volume to 50  $\mu\text{l}/\text{well}$  with Cholesterol Assay Buffer to generate 0, 1, 2, 3, 4, 5  $\mu\text{g}/\text{well}$  of the Cholesterol Standard. (Note: Fluorometric assay is ~10 times more sensitive than the colorimetric assay, the cholesterol standards should be diluted 10 times further if using fluorometric assay).

For sample testing, using 1 to 20  $\mu\text{l}$  of the HDL or LDL/VLDL fraction, adjust the total volume to 50  $\mu\text{l}/\text{well}$  with Cholesterol Assay Buffer. For unknown samples, we suggest testing different volumes of sample to ensure the readings are within the linear portion of the standard curve.

3. Reaction Mix Preparation: Mix enough reagent for the number of assays performed. For each assay, prepare a total 50  $\mu\text{l}$  Reaction Mix containing:

Cholesterol Assay Buffer	44 $\mu\text{l}$
Cholesterol Probe1	2 $\mu\text{l}$
Enzyme Mix	2 $\mu\text{l}$
Cholesterol Esterase2,3	2 $\mu\text{l}$

Notes:

1. If using fluorometric assay, use 0.4  $\mu\text{l}$  of Cholesterol Probe per reaction to reduce fluorescence background.
2. Cholesterol Esterase hydrolyzes cholesteryl ester to free cholesterol. If you want to detect free cholesterol only, omit the Cholesterol Esterase in the reaction, and substitute with 2  $\mu\text{l}$  of Assay Buffer. With the addition of Cholesterol Esterase, the assay detects total cholesterol (cholesterol and cholesteryl esters). If you want to detect cholesteryl esters only, subtract the value of free cholesterol from the value of total cholesterol. Cholesterol Esterase must be added to the reaction for standard curve to convert all the cholesterol in the standard solution.
4. Add 50  $\mu\text{l}$  of the Reaction Mix to each well containing the Cholesterol Standard or test samples, mix well.
5. Incubate the reaction for 60 min at 37°C, protect from light. Measure O.D. at 570 nm or fluorescence at Ex/Em 538/587 nm in a micro-titer plate reader.
6. Calculations: Subtract 0 standard reading from readings. Plot the standard curve. Apply the sample readings to the standard curve to determine sample cholesterol amount in the reaction well. Sample cholesterol concentrations:

$$C = A/V \text{ (}\mu\text{g}/\mu\text{l)}$$

Where: A is the sample cholesterol amount from the standard curve (in  $\mu\text{g}$ ).

V is original sample volume added to the sample reaction well (in  $\mu\text{l}$ ).

Cholesterol Molecular Weight: 386.6; 1  $\mu\text{g}/\mu\text{l}$  = 100 mg/dL.

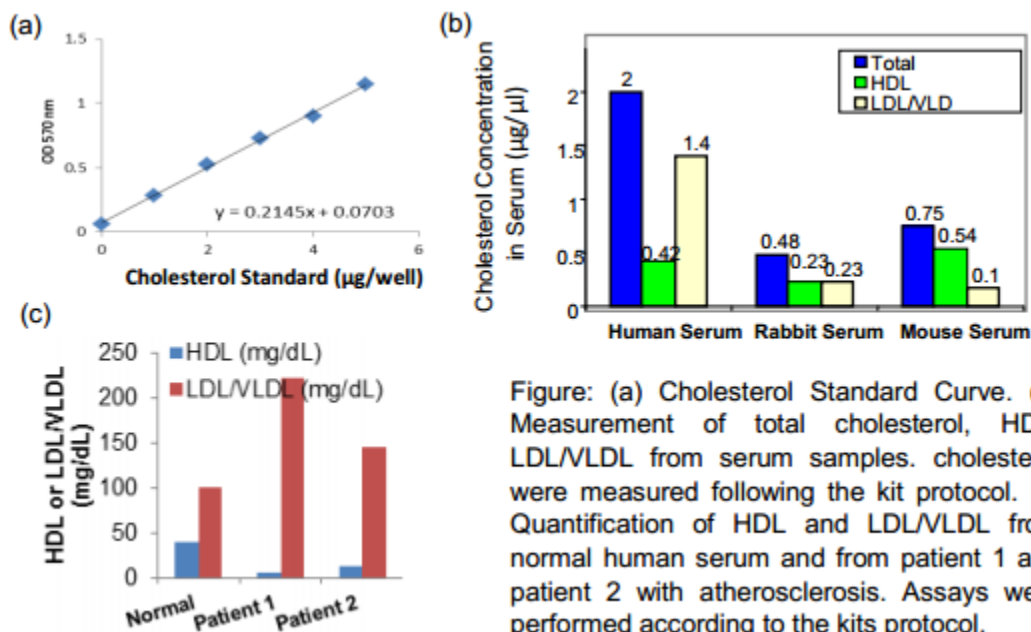


Figure: (a) Cholesterol Standard Curve. (b) Measurement of total cholesterol, HDL, LDL/VLDL from serum samples. cholesterol were measured following the kit protocol. (c) Quantification of HDL and LDL/VLDL from normal human serum and from patient 1 and patient 2 with atherosclerosis. Assays were performed according to the kits protocol.

### General Troubleshooting Guide:

Problems	Cause	Solution
Assay not working	<ul style="list-style-type: none"> <li>• Use of a different buffer</li> <li>• Omission of a step in the protocol</li> <li>• Plate read at incorrect wavelength</li> <li>• Use of a different 96-well plate</li> </ul>	<ul style="list-style-type: none"> <li>• Assay buffer must be at room temperature</li> <li>• Refer and follow the data sheet precisely</li> <li>• Check the wavelength in the data sheet and the filter settings of the instrument</li> <li>• Fluorescence: Black plates ; Luminescence: White plates; Colorimeters: Clear plates</li> </ul>
Samples with erratic readings	<ul style="list-style-type: none"> <li>• Use of an incompatible sample type</li> <li>• Samples prepared in a different buffer</li> <li>• Samples were not deproteinized (if indicated in datasheet)</li> <li>• Cell/ tissue samples were not completely homogenized</li> <li>• Samples used after multiple free-thaw cycles</li> <li>• Presence of interfering substance in the sample</li> <li>• Use of old or inappropriately stored samples</li> </ul>	<ul style="list-style-type: none"> <li>• Refer data sheet for details about incompatible samples</li> <li>• Use the assay buffer provided in the kit or refer data sheet for instructions</li> <li>• Use the 10 kDa spin cut-off filter or PCA precipitation as indicated</li> <li>• Use Dounce homogenizer (increase the number of strokes); observe for lysis under microscope</li> <li>• Aliquot and freeze samples if needed to use multiple times</li> <li>• Troubleshoot if needed, deproteinize samples</li> <li>• Use fresh samples or store at correct temperatures till use</li> </ul>
Lower/ Higher readings in Samples and Standards	<ul style="list-style-type: none"> <li>• Improperly thawed components</li> <li>• Use of expired kit or improperly stored reagents</li> <li>• Allowing the reagents to sit for extended times on ice</li> <li>• Incorrect incubation times or temperatures</li> <li>• Incorrect volumes used</li> </ul>	<ul style="list-style-type: none"> <li>• Thaw all components completely and mix gently before use</li> <li>• Always check the expiry date and store the components appropriately</li> <li>• Always thaw and prepare fresh reaction mix before use</li> <li>• Refer data sheet &amp; verify correct incubation times and temperatures</li> <li>• Use calibrated pipettes and aliquot correctly</li> </ul>
Readings do not follow a linear pattern for Standard curve	<ul style="list-style-type: none"> <li>• Use of partially thawed components</li> <li>• Pipetting errors in the standard</li> <li>• Pipetting errors in the reaction mix</li> <li>• Air bubbles formed in well</li> <li>• Standard stock is at an incorrect concentration</li> <li>• Calculation errors</li> <li>• Substituting reagents from older kits/ lots</li> </ul>	<ul style="list-style-type: none"> <li>• Thaw and resuspend all components before preparing the reaction mix</li> <li>• Avoid pipetting small volumes</li> <li>• Prepare a master reaction mix whenever possible</li> <li>• Pipette gently against the wall of the tubes</li> <li>• Always refer the dilutions in the data sheet</li> <li>• Recheck calculations after referring the data sheet</li> <li>• Use fresh components from the same kit</li> </ul>
Unanticipated results	<ul style="list-style-type: none"> <li>• Measured at incorrect wavelength</li> <li>• Samples contain interfering substances</li> <li>• Use of incompatible sample type</li> <li>• Sample readings above/below the linear range</li> </ul>	<ul style="list-style-type: none"> <li>• Check the equipment and the filter setting</li> <li>• Troubleshoot if it interferes with the kit</li> <li>• Refer data sheet to check if sample is compatible with the kit or optimization is needed</li> <li>• Concentrate/ Dilute sample so as to be in the linear range</li> </ul>

Note: The most probable list of causes is under each problem section. Causes/ Solutions may overlap with other problems.



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