

Proteasome Activity Fluorometric Assay Kit

For detecting proteasome activity in cultured cells using our proprietary green fluorescence probe

For research use only - not intended for diagnostic use.

Storage and Stability

On receipt entire assay kit should be stored at -20°C, protected from light.

Materials Supplied

| Item | Quantity | Storage Condition |
|---|----------|-------------------|
| Component A: Proteasome LLVY-R110 Substrate | 1 vial | -20°C |
| Component B: Assay Buffer | 10 mL | -20°C |
| Component C: DMSO | 100 µL | -20°C |

Δ Note: Thaw all the kit components to room temperature before starting the experiment.

Assay Protocol

Preparation of Cells

For adherent cells: Plate cells overnight in growth medium at 80,000 cells/well/90µL for a 96-well plate or 20,000cells/well/20µL for a 384-well plate.

For non-adherent cells: Centrifuge the cells from the culture medium and then suspend the cell pellet in culture medium at 300,000 cells/well/90µL for a 96-well poly-D lysine plate or 80,000 cells/well/20µL for a 384-well poly-D lysine plate. Centrifuge the plate at 800 rpm for 2 minutes with brake off prior to the experiments.

Δ Note: Note: Each cell line should be evaluated on an individual basis to determine the optimal cell density

Preparation of Proteosome Assay Loading Solution

1. Thaw all the kit components at room temperature before use.
2. Make 400X Proteasome LLVY-R110 Substrate stock solution: Add 25 μ L of DMSO (Component C) to the vial of Proteasome LLVY-R110 Substrate (Component A), and mix well.
3. Make proteasome assay loading solution: Add 25 μ L of 400X Proteasome LLVY-R110 Substrate stock solution (from Step 2) into 10 mL of Assay Buffer (Component B), and mix well.

Δ Note: 25 μ L of 400X Proteasome LLVY-R110 Substrate stock solution (from Step 2) and 10 mL of Assay Buffer (Component B) are enough for 1 plate. Aliquot and store the unused 400X Proteasome LLVY-R110 Substrate stock solution and Assay Buffer at -20 °C. Avoid repeated freeze-thaw cycles.

Run Proteasome Assay:

1. Treat cells with 10 μ L of 10X test compound (for a 96-well plate) or 5 μ L of 5X test compound (for a 384-well plate) in PBS or desired buffer. For blank wells (medium without the cells), add the corresponding amount of compound buffer.
2. Incubate the cell plates in a 5% CO₂, 37 °C incubator for a desired period of time.

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For more details, please visit <http://www.apexbt.com/> or contact our technical team.

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