

## Product Information

### Aspartate Colorimetric/Fluorometric Assay Kit

#### I. Kit Contents:

Components	K2082-100 100 assays	Cap Color	Part Number
Aspartate Assay Buffer	25 ml	WM	K2082-C-1
Probe (DMSO solution)	0.2 ml	Red	K2082-C-2
Serum Clean Up Mix	lyophilized	Blue	K2082-C-3
Aspartate Enzyme Mix	lyophilized	Green	K2082-C-4
Conversion Mix	lyophilized	Purple	K2082-C-5
Aspartate Standard (100 mM)	0.1 ml	Yellow	K2082-C-6

#### II. Introduction:

Aspartate (Asp) is an  $\alpha$ -amino acid. L-Aspartic acid is one of the 23 proteinogenic amino acids and serves as a precursor to four essential amino acids (Met, Thr, Ile and Lys). Aspartate is a metabolite in the urea cycle and is involved in gluconeogenesis and transports reducing equivalents between the mitochondria and the cytosol via the malate-aspartate shuttle. Also, aspartate is an excitotoxin and serves as an excitatory neurotransmitter in the brain through the stimulation of NMDA receptors.

The Aspartate Colorimetric/Fluorometric Assay Kit provides a sensitive, simple and convenient way for detection of aspartate in a variety of samples based on colorimetric and fluorometric method. In the assay, aspartate is converted to pyruvate which is oxidized with the conversion of a probe into a fluorescent (Ex/Em 535/587 nm) and highly colored (570 nm) species proportional to the amount of aspartate in samples. Aspartate can be quantified in the range between 0.1 – 10 nmoles/well (2 - 200  $\mu$ M).

#### III. Reagent Preparation, Storage and Handling:

Store the kit at  $-20^{\circ}$  C prior to use. Read the entire protocol before performing the assay.

Aspartate Probe: Ready to use as supplied. Warm the probe to room temperature to melt the DMSO prior to use.

Serum Clean Up Mix, Aspartate Enzyme Mix, Conversion Mix: Add 220  $\mu$ l of Aspartate Buffer to each vial respectively and dissolve completely prior to use. These can be kept for up to a week after reconstitution. If use over a longer period is anticipated, they should be aliquoted and stored at  $-20^{\circ}$ C.

#### IV. Assay Protocol:

##### 1. Standard Curve Preparation:

Colorimetric Assay: Dilute the Aspartate Standard to 1.0 mM by adding 10  $\mu$ l of the 100 mM Aspartate Standard to 990  $\mu$ l of dH<sub>2</sub>O, mix well. Add 0, 2, 4, 6, 8, 10  $\mu$ l into a series of wells. Adjust volume to 50  $\mu$ l/well with Assay Buffer to generate 0, 2, 4, 6, 8, 10 nmol/well of the Aspartate Standard.

Fluorometric Assay: Dilute the Aspartate standard to 1 mM as in the colorimetric Assay. Dilute further another 10X by taking 100  $\mu$ l of the standard and adding 900  $\mu$ l of dH<sub>2</sub>O mix well. Add 0, 2, 4, 6, 8, 10  $\mu$ l into a series of wells. Adjust volume to 50  $\mu$ l/well with Assay Buffer to generate 0, 0.2, 0.4, 0.6, 0.8, 1.0 nmol/well of the Aspartate Standard.

##### 2. Sample Preparation:

Cell extracts can be used directly in the assay. Serum samples require pretreatment to remove interfering substances: Add 2  $\mu$ l of the Serum Clean Up Mix to 100  $\mu$ l serum and incubate 30 min at room temperature. Treated serum samples should be deproteinized by centrifuging 10 min with a 10 kDa spin filter Filtrate (1 - 30  $\mu$ l) can be used directly in the assay. Adjust all well volumes to 50  $\mu$ l with Assay Buffer. Due to the relatively low levels of aspartate in serum, use of the fluorometric assay is strongly recommended.

### 3. Reaction:

Prepare 50  $\mu$ l of reaction mix for each standard and sample well to be measured. The reaction mix consists of:

	Reaction Mix	Background Control
Aspartate Enzyme Mix	2 $\mu$ l	---
Conversion Mix	2 $\mu$ l	2 $\mu$ l
Probe	2 $\mu$ l	2 $\mu$ l
Aspartate Buffer	44 $\mu$ l	46 $\mu$ l

In order to reduce background in the fluorometric assay, reduce the amount of probe per well to 0.5  $\mu$ l per well

Samples may contain relatively high levels of pyruvate which will increase background. In that case a background control is needed to correct for pyruvate.

4. Incubate: For 30 min at room temperature

5. Read: Measure OD at 570 nm or fluorescence at Ex/Em 535 nm/587 nm in a microplate reader.

### 6. Calculation:

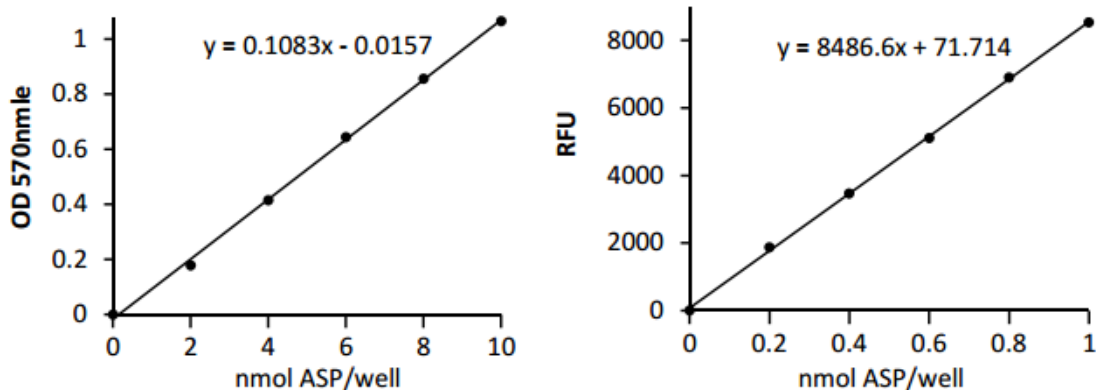
Correct background by subtracting the value derived from the 0 Standard from all readings (The background reading can be significant and must be subtracted). Plot the Standard curve. Read sample concentrations from the standard curve:

$$C = Sa/Sv \text{ nmol}/\mu\text{l or mM,}$$

Where Sa is the sample amount (in nmol) from standard curve.

Sv is the sample volume ( $\mu$ l) added into the wells.

Aspartate MW: 65.384 g/mol.



Colorimetric and Fluorometric standard curves obtained following this protocol.

**For research use only! Not to be used in humans.**



## Our promise

If the product does not perform as described on this datasheet, we will offer a refund or replacement. For more details, please visit <http://www.apexbt.com/> or contact our technical team.

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