Product Information

Calpain Activity Fluorometric Assay Kit

I. Kit Contents:

Components	K2062-100 100 assays	Cap Color	Part Number
Extraction Buffer	25 ml	WM	K2062-C-1
10X Reaction Buffer	1.5 ml	Clear	K2062-C-2
Calpain Substrate Ac-LLY-AFC	0.5 ml	Amber	K2062-C-3
Active Calpain I (Positive Control)	10 µl	Green	K2062-C-4
Calpain Inhibitor Z-LLY-FMK	10 µl	Orange	K2062-C-5

II. Introduction:

Apoptosis is one of the biological processes that activation of calpain is implicated in. Cell membrane and Ca^{2+} is required for calpainactivation. The activated calpain is entered into cytosol. The Calpain Activity Assay Kit is a convenient way of measuring calpain activity with optimized buffers and reagents. The Extraction Buffer extracts cytosolic proteins with contaminations from cell membrane and lysosome proteases. It also protects the cell from atuo-activation of calpain during extraction process. Thus, the kit only detects activated calpain in cytosol following treatment of cells with chemicals or drugs inducers. The fluorometric assay is based on the detection of cleavage of calpain substrate Ac-LLY-AFC. Ac-LLY-AFC emits blue light (λmax = 400 nm); upon cleavage of the substrate by calpain, free AFC emits a yellow-green fluorescence (λmax = 505 nm), which can be quantified using a fluorometer or a fluorescence.

III. Calpain Assay Procedure:

1. Treat cells by desired methods. Concurrently incubate a control culture without treatment.
2. Count cells and pellet ~ 1 - 2 x 10^6 cells by centrifugation.
3. Resuspend cells in 100 µl Extraction Buffer and incubate samples on ice for 20 minutes. Gently mix the samples by tapping several times during incubation.
4. Centrifuge for 1 min in a microcentrifuge (10K x g) and transfer supernatant to a fresh tube and put on ice. Assay protein concentration.
   Note: because if the high reducing agent content in the extration buffer-dilute about 10-fole then use a Coomassie-based protein assay.
5. Dilute the cell lysate (~ 50 - 200 µg) to 85 µl of Extraction Buffer.
   For positive control, add 1 - 2 µl Active Calpain to 85 µl of Extraction Buffer.
   For negative control, use untreated cell lysate or add 1 µl Calpain Inhibitor to the treated cell lysate.
6. Add 10 µl of 10X Reaction Buffer and 5 µl of Calpain Substrate to each assay.
7. Incubate at 37℃ for 1 hour in the dark.
8. Read samples in a fluorometer equipped with a 400 nm excitation filter and 505 nm emission filter. For a plate reading set up, transfer the samples to a 96-well plate.
9. The changes in calpain activity can be determined by comparing results of treated samples and negative control. Alternatively, the activity can be expressed as Relative Fluorescent Unit (RFU) per milligram protein of each sample.

IV. Storage and Stability:

Store kit at -70℃ (Store Extraction Buffer and 10X Reaction Buffer at 4℃ after opening).

All reagents are stable for 6 months under proper storage conditions.

General Troubleshooting Guide:

Problems	Cause	Solution
Assay not working	• Use of ice-cold assay buffer • Omission of a step in the protocol • Plate read at incorrect wavelength • Use of a different 96-well plate	• Assay buffer must be at room temperature • Refer and follow the data sheet precisely • Check the wavelength in the data sheet and the filter settings of the instrument • Fluorescence: Black plates ; Luminescence: White plates; Colorimeters: Clear plates
Samples with erratic readings	• Use of an incompatible sample type • Samples used after multiple free-thaw cycles • Presence of interfering substance in the sample • Use of old or inappropriately stored samples	• Refer data sheet for details about incompatible samples • Aliquot and freeze samples if needed to use multiple times • Troubleshoot if needed, deproteinize samples • Use fresh samples or store at correct temperatures till use
Lower/ Higher readings in Samples and Standards	• Improperly thawed components • Use of expired kit or improperly stored reagents • Allowing the reagents to sit for extended times on ice • Incorrect incubation times or temperatures • Incorrect volumes used	• Thaw all components completely and mix gently before use • Always check the expiry date and store the components appropriately • Always thaw and prepare fresh reaction mix before use • Refer data sheet & verify correct incubation times and temperatures • Use calibrated pipettes and aliquot correctly
Readings do not follow a linear pattern for Standard curve	• Use of partially thawed components • Pipetting errors in the standard • Pipetting errors in the reaction mix • Air bubbles formed in well • Standard stock is at an incorrect concentration • Calculation errors • Substituting reagents from older kits/ lots	• Thaw and re-suspend all components before preparing the reaction mix • Avoid pipetting small volumes • Prepare a master reaction mix whenever possible • Pipette gently against the wall of the tubes • Always refer the dilutions in the data sheet • Use fresh components from the same kit • Recheck calculations after referring the data sheet
Unanticipated results	• Measured at incorrect wavelength • Samples contain interfering substances • Use of incompatible sample type • Sample readings above/below the linear range	• Check the equipment and the filter setting • Troubleshoot if it interferes with the kit • Refer data sheet to check if sample is compatible with the kit or optimization is needed • Concentrate/ Dilute sample so as to be in the linear range
Note: The most probable list of causes is under each problem section. Causes/ Solutions may overlap with other problems.		

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Our promise

If the product does not perform as described on this datasheet, we will offer a refund or replacement. For more details, please visit http://www.apexbt.com/ or contact our technical team.

Tel: +1-(832)696-8203
Fax: +1-832-641-3177
Email: sales@apexbt.com