

## **Product Information**

# **Apoptotic DNA Ladder Isolation Kit**

#### I. Kit Contents:

Component	K2045-50	Cap Colorr	Part Number
	50 assays		
DNA Ladder Extraction Buffer	12.5 ml	WM	K2045-C-1
Enzyme A Solution	0.25 ml	Blue	K2045-C-2
Enzyme B (Lyophilized)	1 vial	Red	K2045-C-3
Ammonium Acetate Solution	0.25 ml	Yellow	K2045-C-4
DNA Suspension Buffer	2 ml	Green	K2045-C-5

#### **II. Introduction:**

Internucleosomal DNA fragmentation is a sign of apoptosis in mammalian cells. The Apoptotic DNA Ladder Isolation Kit is a simple and sensitive way of detecting DNA fragmentation in apoptotic cells. It can selectively isolates DNA ladders and will not effected by intact genomic DNA, in turn it prominently increase the cell numbers that can be extracted and load on a single well of agarose gel and thus increase the detection sensitivity.

### **III. Reagent Preparation:**

Dissolve Enzyme B with 275 µl ddH<sub>2</sub>O and mix well before use. The Enzyme B solution should be aliquoted and freeze at -70°C immediately.

#### IV. DNA Ladder Detection Protocol:

- 1. Induce apoptosis in cells by desired method. Concurrently incubate a control culture without induction.
- 2. Wash cells with PBS (not provided) and pellet 2 x 10<sup>6</sup> cells by centrifugation for 5 min at 500 x g. Carefully remove supernatant using pipette. For adherent cells, gently trypsinize cells and then pellet cells.

For tissue samples, cut 50 mg tissues into very fine pieces or homogenize tissues in PBS to generate cell suspension (Note: do not sonicate). Centrifuge to collect cell pellet.

Note: The kit can detect DNA ladder from  $10^5$  apoptotic cells (100% apoptosis). However, if the level of apoptosis in your sample is low, you can increase the cell number up to  $10^7$ . If using more than  $2 \times 10^6$  cells per assay, you should proportionally increase the volume of all reagents.

- 3. Extract the cell pellet with 50  $\mu$ l DNA Ladder Extraction Buffer for 10 seconds at room temperature with gentle pipetting. Centrifuge for 5 min at 1600 x g (~4500 rpm). Transfer the supernatant to a fresh tube.
- 4. Extract the pellet again by repeating step 3. Combine the supernatant.
- 5. Add 5 μl Enzyme A Solution into the supernatant, mix by gentle vortex and incubate at 37 °C for 10 min. (Note: If cells contain high level of DNase, then the incubation step should be skipped, as high level DNase can digest DNA ladder generating smear pattern.)
- 6. Add 5 µl Enzyme B Solution into each sample and incubate at 50 °C for 30 min or longer (overnight is ok).
- 7. Add 5 μl Ammonium Acetate Solution to each sample and mix well. Add 100 μl isopropanol (not provided), mix well, and keep at -20°C for 10 minutes
- 8. Centrifuge the sample at maximum speed ( $\sim$ 16K x g) for 10 minutes to precipitate DNA. (Note: Microcentrifuges typically generate  $\sim$  16K x g at 13K x rpm)



- 9. Remove supernatant, wash the DNA pellet with 0.5 ml 70% ethanol, centrifuge again at maximum speed
- (~16K x g) to remove trace ethanol, and air dry for 10 minutes at room temperature.
- 10. Dissolve the DNA pellet in 30 µl DNA Suspension Buffer (Note: No other loading buffer needed for loading to the gel).
- 11. Load 15-30 µl of the sample onto a 1.2% agarose gel containing 0.5 µg/ml ethidium bromide in both gel and running buffer.
- 12. Run the gel at 5 V/cm for 1-2 hours or until the yellow dye (included in the suspension buffer) run to the edge of the gel.
- 13. Ethidium bromide-stained DNA can be visualized by trans-illumination with uv light and photographed.

For research use only! Not to be used in humans.

## Our promise

If the product does not perform as described on this datasheet, we will offer a refund or replacement. For more details, please visit <a href="http://www.apexbt.com/">http://www.apexbt.com/</a> or contact our technical team.

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