M-NFS-60 Cells

Product information

Cell line	M-NFS-60
Alias	NFS-60; NFS 60; NFS60
Organism	Mouse
Product format	1 x 10^6 Cells/Frozen vial
Cell morphology	lymphoblast
Growth properties	Suspension growth
Culture medium	RPMI-1640+0.05 mM β-mercaptoethanol+62 ng/mL M-CSF+10%FBS+1% P/S
Culture conditions	95% air, 5% CO_2, 37°C
Passage ratio	1:2-1:3
Cryopreservation	Serum-Free Cell Freezing Medium
Cell description	M-NFS-60 cells are isolated from mouse myeloid leukemia and are commonly used in the study of leukemia and immune system diseases.

Protocol

1. Cell reception

1) Upon receipt, please check the package. Take photos and contact us within 3 days if any abnormalities such as dry ice has ran out, the vial is broken or thawed.

2) Directly proceed to cell thawing, or transfer the vial to liquid nitrogen or store briefly at -80°C freezer.

3) Quickly thaw the vial in a 37°C water bath with shaking. Keep the cap out of the water to avoid cell contamination. When the cells are almost thawed (only a little ice crystal remains), stop the water bath, and continue to shake the vial until the ice crystal melts.

4) Wipe the surface of the vial with 75% alcohol. Carefully transfer cells to centrifuge tubes containing 5 mL of pre-warmed complete medium. Centrifuge cells at 1000 rpm for 5 min.

5) Discard the supernatant, resuspend cells with 4-6 mL of complete medium, and seed in a T25 vial (or 6 cm plate).

6) Culture overnight, change the medium the next day. After that, change the medium every 2-3 days.

***Note:** If the cells are in poor condition after thawing, contact us in time and we can resend the product once for free.

2. **Cell passage**

1) Perform the cell passage by the addition of fresh medium or replacement of medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension at 2-4×10^5 cells/mL.

2) Change fresh medium every 2-3 days depending on the cell density.

3. **Cell cryopreservation**

1) Transfer the cell suspension to a tube. Centrifuge at 1000 rpm for 5 min.

2) Discard the supernatant, gently resuspend the cells in an appropriate volume of Cell Freezing Medium at a concentration of 5x10^6-1x10^7 cells/mL.

3) Aliquot the cell suspension into labeled sterile cryovials and tighten the cap of the cryovials.

***Note:** Always use sterile cryovials to store frozen cells. Do not use centrifugal tubes instead of cryovials. Centrifugal tubes don't reliably seal and may fill with liquid nitrogen when frozen, leading them to burst on thawing.

4) Transfer the cryovials into a cell freezing container and immediately store the container at -80ºC for a least 24 h. Then the frozen cells should be transferred to the vapor phase of liquid nitrogen for long-term storage.

***Note:** The cryovials in the cell freezing container should be transferred to -80ºC as soon as possible.

Notes

1. For your safety and health, please wear lab coats and gloves during the experiment.
2. For research use only. Not to be used in clinical diagnostic or clinical trials.

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