

APENBI

ER-Tracker Green

Introduction

ER-Tracker Green is a green-fluorescent endoplasmic reticulum (ER) probe that selectively labels ER of live cells. ER-Tracker Green is the BODIPY FL labeled glibenclamide. Glibenclamide, a drug for diabetic patients, specially binds to the sulphonylurea receptors of ATP-sensitive K⁺ channels on ER. So fluorescent-labeled glibenclamide (ER-Tracker Green) can label ER. Unlike conventional ER probe DiOC₆(3), ER-Tracker Green does not appear to be toxic to cells at low concentrations, while it is highly selective for ER and rarely stains mitochondria. ER-Tracker Green stains live cells well but is not well-retained after fixation with formaldehyde. Glibenclamide may affect ER function, and variable expression of sulphonylurea receptors in some special cell types may lead to non-ER labeling.

This product is supplied as a lyophilized powder. Dissolve a vial in 20 µL of DMSO to make 1 mM ER-Tracker Green Stock Solution.

Components and Storage

	R8812.16 ug
Components	воо 12-10 µу
ER-Tracker Green	16 µg
This product should be stored at -20°C away from light and moisture, stable for at least 6 months.	

Properties

Physical Appearance	Lyophilized powder
M.Wt	783.09
Cas No.	730931-46-1
Formula	C ₃₇ H ₄₂ BCIF ₂ N ₆ O ₆ S
Ex/Em	504/511
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Protocol

 Preparation of the stock solution: Allow ER-Tracker Green to warm to room temperature before use, then briefly centrifuge to deposit the lyophilized powder at the bottom of the vial. Dissolve a vial in 20 µL of DMSO to make 1 mM ER-Tracker Green Stock Solution. Unused ER-Tracker Green Solution (1 mM) should be stored at -20°C away from light and moisture. 2. Preparation of the working solution: Dilute appropriate ER-Tracker Green (1 mM) in a suitable buffer (for example, HBSS with Calcium and Magnesium) to make a working solution. The recommended concentration of the working solution is 1 µM. To reduce potential labeling artifacts, keep the concentration of working solution as low as possible. It is suggested to dilute ER-Tracker Green when using it.

*Note: The optimal concentration of working solution varies depending on the type of cells, the recommended ratio of dilution is 1:1000-1:3000.

3. Labeling of ER: For adherent cells, grow cells to reach the desired density. Remove the growth medium and wash with a suitable buffer (for example, HBSS with Calcium and Magnesium). Add a pre-warmed (37°C) working solution to cover the cells. Incubate at 37°C away from light for 15-30 min. Replace the working solution with a fresh medium. Then detect the fluorescence signal of cells by a microscope with a FITC filter set.

*Note: The optimal time for incubation varies depending on the type of cells. For suspension cells, harvest cells and perform similarly to the adherent cells.

4. Fixation of cells (optional step): If fixation is needed, fix cells with 4% formaldehyde for 2 min at 37°C. after fixation, wash cells 2 times in a suitable buffer prior to further staining, mounting and viewing.

*Note: Permeabilization is not recommended. Permeabilization with Triton X-100 results in no fluorescence signal.

Note

- 1. Fluorescent probes are easy to quench, please protect them from light when using.
- 2. For your safety and health, please wear lab coats and gloves during the experiment.
- 3. For research use only. Not to be used in clinical diagnostic or clinical trials.

